

# Constraints on Metabolic Network Analysis in Bacterial Physiology

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# Introduction

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PRX Life PRX Life 3, 022001 Published 1 April 2025 [9].

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Main ideas of this paper are:

- Characterize **emergent properties** of biological interactions in bacterial cells.
- These constraints are equivalent to **Kirchhoff's laws** and **Ohm's law**.
- Bacterial growth physiology can be analyzed quantitatively as **electrical circuits**  $\Rightarrow$  coarse-graining.<sup>1</sup>

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## Growth Laws

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## Growth Laws

Many emergent behaviors can be described by simple phenomenological laws:

- (i) Rate at which environmental materials are assimilated is balanced according to composition
- (ii) Rates are constrained by the autocatalytic nature of life

- When environmental nutrient is unlimited, population increases like

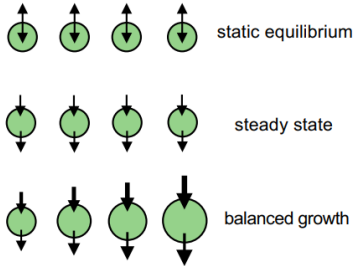
$$\frac{dN}{dt} \sim N(t) \Rightarrow N(t) = N_0 e^{\lambda t}. \quad (1)$$

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- **Balanced growth** characterizes exponential phase: In order for cells to accumulate exponentially, generating processes must happen at balanced rates.

# Metabolic Networks Are Complicated



**Figure 1:** Comparison of equilibrium, steady state, and balanced growth [6].

# Metabolic Networks Are Complicated

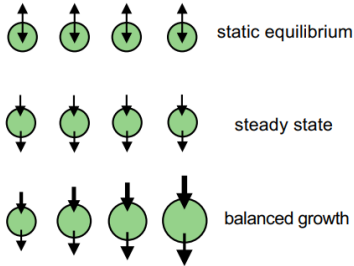


Figure 1: Comparison of equilibrium, steady state, and balanced growth [6].

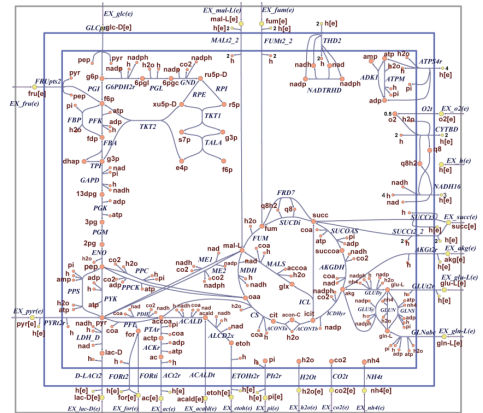


Figure 2: Core metabolic network of E. coli [7].

- Stoichiometric matrix  $S \in M_{m \times n}(\mathbb{R})$ , biomass vector  $X \in \mathbb{R}^n$ :

$$\frac{dX}{dt} \equiv J = SX \quad (2)$$

$S$  is *underspecified* (metabolism is an open system) and *sparse*.

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- Evolution selects cells that grow fast:  $Z \propto \lambda$
- Metabolic reaction rates must be **balanced** during steady-state<sup>2</sup> growth:  $J = 0$ .

---

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# Flux Balance is a Linear Programming Problem

## Constrained Optimization

Maximize the objective function  $Z = c \cdot x$   
subject to

$$J = Sx = 0 \quad (\text{balanced growth}), \quad (3)$$

and

$$lb_i \leq x_i \leq ub_i \quad (\text{bounded rates}). \quad (4)$$

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## Proteome Partition of E. coli

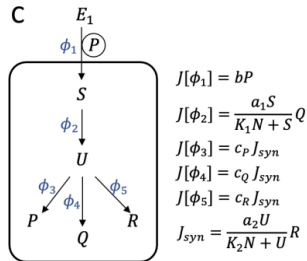


Figure 3: Three-sector proteome partition model (Lin, Wei-Hsiang, 2025) [9, 5]

# Ribosomes Catalyze Protein Synthesis

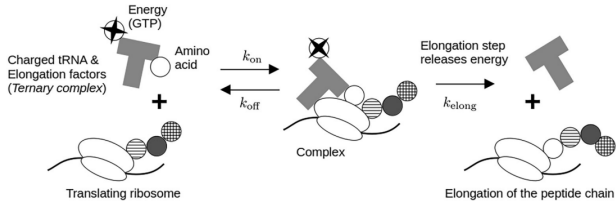
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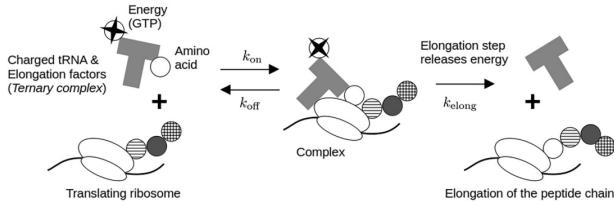
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**Figure 4:** Ribosomes follow similar kinetics to those of enzymes: they turn charged tRNA into uncharged tRNA.

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**Figure 4:** Ribosomes follow similar kinetics to those of enzymes: they turn charged tRNA into uncharged tRNA.

(Haldane) Abundance of the substrate far exceeds the abundance of the enzyme.

$$\text{rate} \propto [\text{Rb}] \times \frac{[\text{tRNA}]}{K_M + [\text{tRNA}]} \quad (5)$$





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Protein mass fraction is a **linear** function of growth rate:

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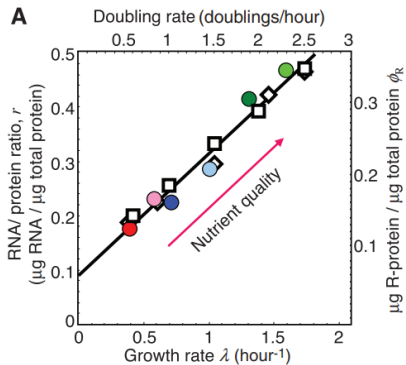
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Ohm's law:  $\Delta V = I/G$ .

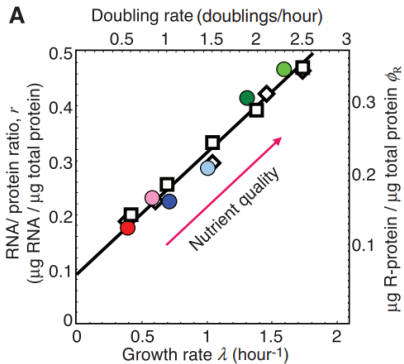


# Global Constraints

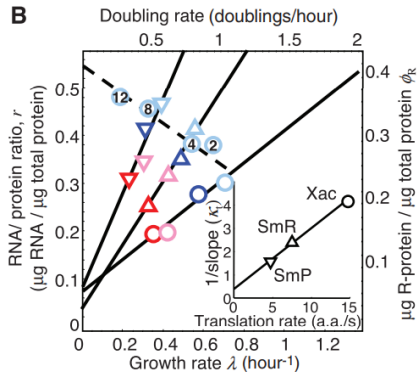


**Figure 5:** Growth rate is modulated by quality of nutrient [8].

# Global Constraints



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**Figure 6:** Growth rate is modulated by translational inhibition [8].



## Proteomic Coarse-Graining and Electric Circuit

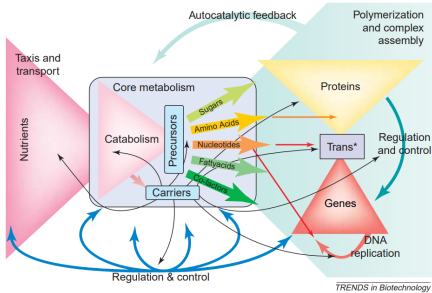
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Conserved large-scale topological features among micro-organisms:

# Bow-Tie Topology

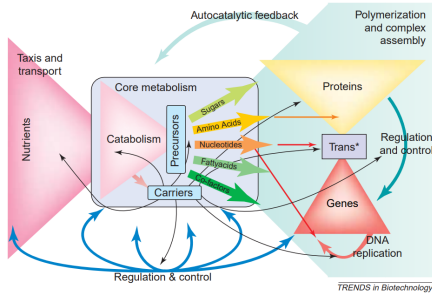
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**Figure 7:** Common module for bacterial metabolism: diversity of inputs and outputs, processed with few intermediate common currencies [3].

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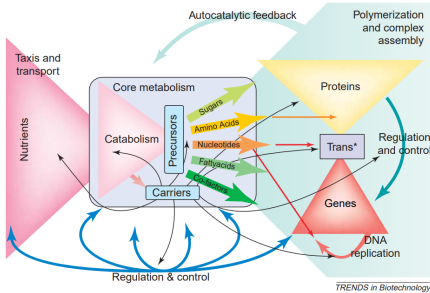


- Bacterial metabolism and transcriptional machinery exhibits **bow tie architecture**

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# Bow-Tie Topology

Conserved large-scale topological features among micro-organisms:



- Bacterial metabolism and transcriptional machinery exhibits **bow tie architecture**
- Proteins can be partitioned into only few classes

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# Equivalent Circuits and Kirchhoff's Laws

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## Kirchhoff's Laws

Governing laws for DC circuits

$$\sum_{\text{node } m} j_n = 0 \text{ (current law),} \quad (7)$$

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## Kirchhoff's Laws

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## Thevenin's Law

A network of voltage sources and resistors can be replaced by an equivalent circuit with one voltage source and one resistor.

# Anti-Correlation Among Proteome Sectors

- **Proteome partition**: coarse-graining proteins into sectors that behave similarly under specific probes, e.g. functionality

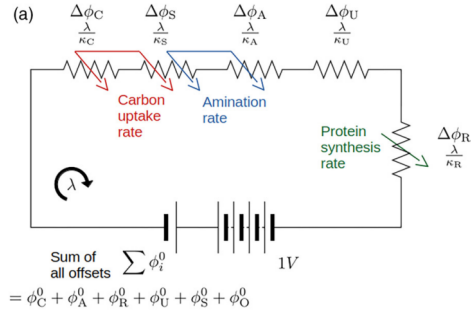
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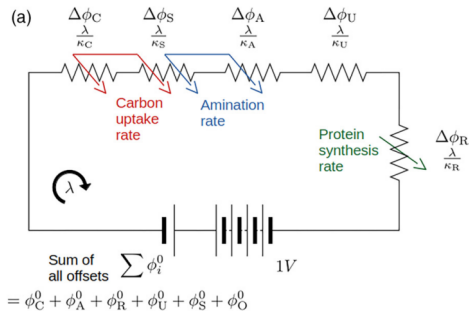
- **Proteome partition**: coarse-graining proteins into sectors that behave similarly under specific probes, e.g. functionality
- Two sectors:  $\phi_M^0 + \phi_R^0 = 1$
- Antibiotic decreases  $\lambda$  without affecting  $\phi_M^0$ : modulates  $\kappa_R$  alone.
- Nutrient quality modulates  $\kappa_M$ .

# Ohmics



**Figure 8:** Six-sectors: ribosomes (R), carbon uptake (C), a.a. biosynthesis (A), carbon uptake + a.a. biosynthesis (S),  $\lambda$ -dependent but not inhibited (U), not  $\lambda$ -dependent [4].

# Ohmics

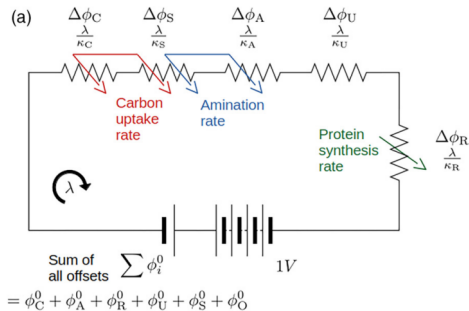


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Coarse-grain according to proteins' response to probes.

$$\lambda = \frac{1 - \phi_C^0 - \phi_A^0 - \phi_R^0 - \phi_U^0 - \phi_S^0 - \phi_O^0}{1/\kappa_C + 1/\kappa_A + 1/\kappa_R + 1/\kappa_U + 1/\kappa_S} \quad (9)$$

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Growth on  $N$  carbon sources:

$$\frac{1}{\kappa_C} \rightarrow \frac{1}{\kappa_{C_1} + \kappa_{C_2} + \dots + \kappa_{C_N}} \quad (10)$$

# Applications

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- Ribosome-targeting antibiotics can modulate conductance  $\kappa_R$ .

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- "Ohmics" assumption for antibiotics-growth rate relationship:

$$\begin{cases} \frac{da}{dt} &= -\lambda a - k_{\text{on}} a r_u + k_{\text{off}} r_b + P_{\text{in}} a_{\text{ex}} - P_{\text{out}} a, \\ \frac{dr_u}{dt} &= -\lambda r_u - k_{\text{on}} a r_u + k_{\text{off}} r_b + s(\lambda), \\ \frac{dr_b}{dt} &= -\lambda r_b + k_{\text{on}} a r_u - k_{\text{off}} r_b. \end{cases} \quad (11)$$

$a_{\text{ex}}, k_{\text{on}}, k_{\text{off}}, P_{\text{in}}, P_{\text{out}} \in \mathbb{R}_{\geq 0}$ .  $s(\lambda)$  is undetermined!

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- Qualitatively different behavior based on binding affinity.

# Antibiotic Transport and Binding

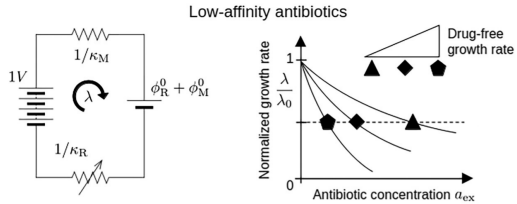


Figure 9: Low-affinity antibiotics [9].

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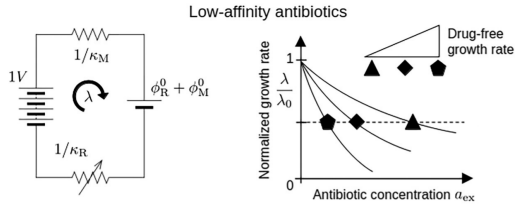


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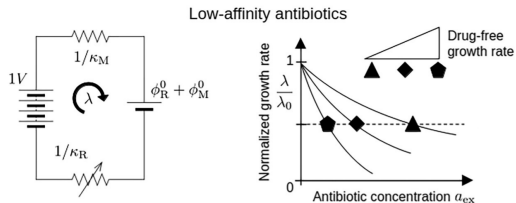


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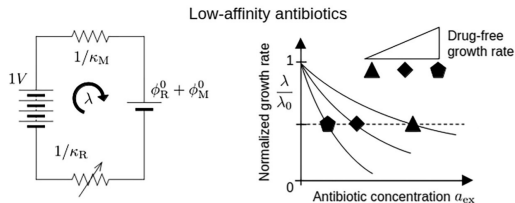


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- Effective against **fast-growing** bacteria.



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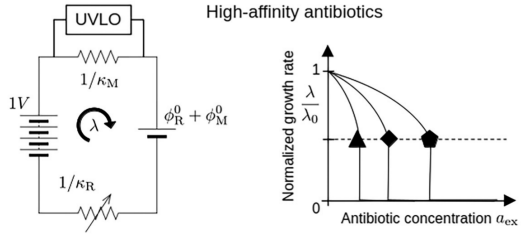


Figure 10: High-affinity antibiotics [9].

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- Sigmoidal inhibition curves.

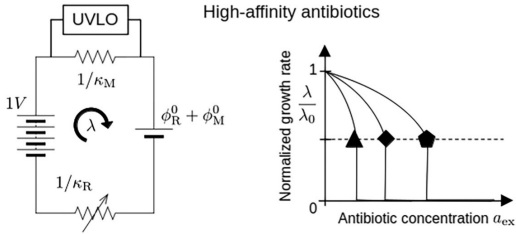


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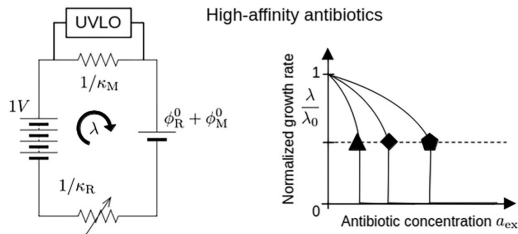


Figure 10: High-affinity antibiotics [9].

- Sigmoidal inhibition curves.
- Half-inhibition conc. correlated with growth rate:

$$\lambda_0 = \lambda \left[ \frac{1}{2} \left( 1 + \sqrt{1 - \frac{a_{\text{ex}}}{IC_{50}}} \right) \right]^{-1}.$$

- Abrupt drop of  $\lambda$  at  $IC_{50}$  analogous to an undervoltage lockout (UVLO)

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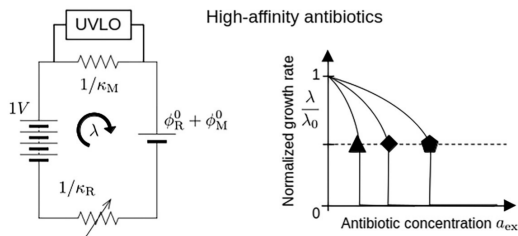


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- Abrupt drop of  $\lambda$  at  $IC_{50}$  analogous to an undervoltage lockout (UVLO)
- Effective against **slow-growing** bacteria.



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## Adapting *E. coli* to growth in glucose and citrate [1]

- Parameters unchanged except for decrease in  $\phi_O^0, \phi_A^0, \phi_S^0$ .

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## Adapting *E. coli* to growth in glucose and citrate [1]

- Parameters unchanged except for decrease in  $\phi_O^0, \phi_A^0, \phi_S^0$ .
- Mechanistic explanation:
  - $\phi_O^0$ : decrease in porin *OmpF*
  - $\phi_A^0, \phi_S^0$ : enzymes associates with pyruvate kinase *PykF*.



## Concluding Remarks

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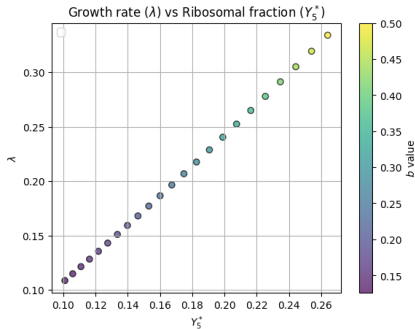


Figure 11: Three-sector partition (low nutrient).

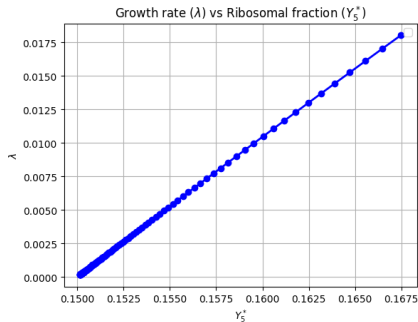


Figure 12: Six-sector partition (low nutrient).

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- Mechanistic justification for coarse-graining complex biochemical networks with circuits.
- Wealth of large-Ohmics data → opportunity for synthetic biology.



Thank You  
Q & A

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